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74) Agent: LAMMERT, Steven, R.; Barnes & Thornburg, 11 South Meridian Street, Indianapolis, IN 46204 (US). 54) Title: REGULATORY ELEMENT FOR EXPRESSING GENES IN PLANTS 57) Abstract The present invention is directed to a novel promoter sequence for expressing genes in eukaryotic cells and the construction	(22) International Filing Date: 3 April 1998 (c) (30) Priority Data: 60/042,926 4 April 1997 (04.04.97) (71) Applicant (for all designated States except US): For RESEARCH FOUNDATION [US/US]; Room 30 Hovde Hall, West Lafayette, IN 47907–1021 (US) (72) Inventors; and (75) Inventors/Applicants (for US only): HODGES, The [US/US]; 6038 West 75 N., West Lafayette, IN 479 LYZNIK, Leszek, A. [PL/US]; 5960 N.W. 62nd	03.04.99 UPURDU 07, 102).	BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, G GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, K LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MV MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, S TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIP patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasis patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), Europee patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GI IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, C CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG). Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt
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REGULATORY ELEMENT FOR EXPRESSING GENES IN PLANTS

Field of the Invention

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The present invention is directed to nucleic acid sequences that control the expression of genes in eukaryotic cells. More particularly, the invention is directed to a gene promoter that confers a high level of expression to genes that are operably linked to the promoter.

Background and Summary of the Invention

The present invention relates to a novel regulatory element which confers a high level of expression in plant cells to genes that are operably linked to the regulatory element. The ability to control the level of gene expression in plants is important for many applications of genetic transformation procedures including those directed to crop improvement.

In eukaryotic organisms, multi-level regulatory systems exist to control gene expression. The transcription process is an integral part of such systems and is involved in synthesis of mRNA molecules. The efficiency of transcription is mostly determined by a region of DNA called the promoter. The promoter consists of gene sequences upstream of the site of transcription initiation. The components of the promoter region include the "TATA" box and often a "CAAT" box. In addition, many other regulatory elements that affect transcription may be present in the promoter sequences. The coordinated action of cellular proteins (transcription factors) interacting with promoter sequences determines the specificity of a particular promoter and its effectiveness. Since most eukaryotic genes are stringently regulated, there is a limited availability of promoters with constitutive, strong expression.

The present invention describes the isolation and purification of a DNA sequence that expresses operably linked genes to high levels in plant cells. The promoter sequence described in the present invention expresses genes at a level equal to or higher than that obtained from one of the strongest presently available promoters - the 35S cauliflower mosaic virus promoter. Such promoters are needed to direct a high level of protein expression in transgenic plants. The strong promoter of the present invention is used to construct expression vectors for expressing genes in plant

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cells. In one embodiment, a plant expression vector is provided that comprises the regulatory element of SEQ ID NO: 2 operably linked to a non-natively associated gene, and this vector is used to produce transgenic plants.

5 Brief Description of the Drawings

Fig. 1 Represents a restriction map of the 4.8 kb *Hind*III *Arabidopsis* genomic fragment that hybridizes to the *BgI*I RTS-1 gene fragment.

Fig. 2 Expression of gusA in Arabidopsis protoplasts when gusA is operably linked to: the 35S cauliflower mosaic virus promoter (p35GUS), the promoter of SEQ ID NO: 2 (pUN-GUS), the promoter of SEQ ID NO: 3 (pASR-GUS) or lacking a promoter (DNA-).

Detailed Description of the Invention

Definitions

Unless specified otherwise, any reference to DNA, a DNA sequence, promoter, or regulatory sequence is a reference to a double stranded DNA sequence. A promoter is a DNA sequence that directs the transcription of a structural gene. Typically, a promoter is located in the 5' region of a gene, proximal to the transcription start site of a structural gene. If a promoter is an inducible promoter, then the rate of transcription increases in response to an inducing agent. In contrast, if the promoter is a constitutive promoter, then the rate of transcription is not regulated by an inducing agent.

An enhancer is a DNA regulatory element that can increase the efficiency of transcription, regardless of the distance or orientation of the enhancer relative to the start site of transcription.

The term "expression" refers to the biosynthesis of a gene product. For example, in the case of a structural gene, expression involves the transcription of the structural gene into messenger RNA and the translation of messenger RNA into one or more polypeptides.

An expression vector is a DNA molecule comprising the regulatory elements necessary for transcription of a gene in a host cell. Typically the gene is placed under the control of certain regulatory elements including constitutive or

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inducible promoters, tissue-specific regulatory elements, and enhancer elements. Such a gene is said to be "operably linked to" the regulatory elements when the regulating element controls the expression of the gene. Expression vectors typically include eukaryotic and/or bacterial selectable markers that allow for selection of cells containing the expression vector.

An exogenous DNA sequence refers to a DNA sequence that has been introduced into a host cell from an external source. A transgenic plant is a plant having one or more plant cells that contain an exogenous DNA sequence. The term stably transformed refers to a transformed cell or plant that is capable of transmitting an exogenous DNA sequence to its progeny. Typically a stably transformed host has the exogenous DNA sequence integrated into its genome.

A core promoter contains the essential nucleotide sequences for promoter function, including the TATA box and start of transcription. By this definition, a core promoter may or may not have detectible activity in the absence of specific sequences (regulatory elements) that may enhance the activity of the core promoter or confer tissue specific activity.

A visible marker is defined herein as including any gene that encodes a product that produces a phenotypic trait to the host cell or organism.

A selectable marker is defined herein as including any nucleic acid sequence or gene product that can be selected for after introduction into a cell. The selectable marker facilitates the identification of transformants.

A polylinker is a DNA sequence that contains multiple endonuclease restriction enzyme identification sequences in close proximity of one another.

The present invention is directed to a substantially purified genomic

DNA sequence isolated from *Arabidopsis thaliana* (SEQ ID NO: 1). The genomic

DNA encodes for two proteins (ASR-2 and ORF 3) and contains a dual promoter

region located between those two genes that drives the expression of both genes (see

Fig. 1).

The genomic region containing the coding DNA sequence for ASR-2,

located between nucleotides 945 to 3694 of SEQ ID NO: 1, encompasses sequences
that are homologous to a human pre-mRNA splicing factor ASF/SF2 and the

Arabidopsis SR1 gene. The alignment of the ASR-2 genomic DNA sequences with

the SR1 cDNA sequence indicated the presence of eleven putative exons in the ASR-2 gene, and the deduced amino acid sequence has 82% identity (92% similarity) with the deduced amino acid sequence of SR1. The sequence identity of ASR-2 with the human splicing factor SF2 was 62% as compared to 59% identity between the SR1 and SF2 genes. The ASR-2 gene also appears to have an identical structural organization of RNA-binding domains, the glycine spacer, and the SR domain as is observed in the SR1 and SR2 genes. The ASR-2 coding sequence also includes a highly charged PSK domain at the C-terminal end similar to the SR1 gene but absent in the ASF/SF-2 gene coding sequences.

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The regulatory elements controlling the expression of the ASR-2 gene are contained within the 530 nucleotide region shown as SEQ ID NO: 3. The expression of the ASR-2 gene was analyzed by reverse transcription PCR in different parts of *Arabidopsis* plant. The ASR-2 gene was found to be expressed in all plant parts investigated including the leaves, stems, siliques, and roots. Similar levels of expression were observed in different plant organs. The experiment revealed the presence of more than one transcript hybridizing to the ASR-1 probe (the 2.4 kb *Eco*RI fragment of ASR-1 genomic clone), and could represent splice variants of ASR-2 transcripts.

The 4.8 kb HindIII genomic fragment encodes for another gene, ORF 3 that is located on the complimentary strand relative to the sequence encoding the ASR-2 gene (see Fig. 1), between nucleotides 4217 and 4917 of SEQ ID NO: 1. A 530 bp region is located between the ASR-2 and ORF 3 genes (at position 3691-4220 of SEQ ID NO: 1) and that 530 bp region functions as a dual promoter for expressing both ASR-2 and ORF 3. The sequence of the DNA region that contains the regulatory elements for expressing ORF 3 is shown as SEQ ID NO: 2.

SEQ ID NO: 2 and SEQ ID NO: 3 are inverse compliments of each other, and accordingly a double stranded DNA sequences that contains SEQ ID NO: 2 will also contain SEQ ID NO: 3. As used herein with reference to double stranded DNA sequences, SEQ ID NO: 2 and SEQ ID NO: 3 will designate the orientation of the 530 bp region in DNA constructs. If the 530 bp region is ligated to a gene through its 3' end (as shown in SEQ ID NO: 1), the sequence will be referred to as SEQ ID NO: 2, and if the 530 bp region is ligated to a gene through its 5' end, the sequence

will be referred to as SEQ ID NO: 3. For example, a gene operably linked to a promoter comprising the sequence of SEQ ID NO: 2 designates that the promoter is operably linked to that gene in the orientation naturally expresses the ORF 3 gene.

The 530 bp region located between the ASR-2 and the ORF 3 genes promoter region contains sequences that are known to bind proteins that are involved in the transcriptional process and can function in either direction. Accordingly, this sequence can be used in either orientation as a promoter for expressing genes in eukaryotic cells and more particularly in plant cells. The present invention is directed to a substantially pure DNA sequence comprising the sequence of SEQ ID NO: 2, and the use of such a sequence to express exogenous genes in plants.

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In accordance with one embodiment, a recombinant expression vector is prepared comprising a promoter having a consecutive 20 base pair sequence identical to the sequence of SEQ ID NO: 2 or SEQ ID NO: 3. Typically the expression vector will also include a polylinker region located adjacent to the promoter such that upon insertion of a gene sequence into the polylinker, the gene will be operably linked to the promoter. In one embodiment the promoter utilized is the DNA sequence of SEQ ID NO: 2. The expression vector typically includes a eukaryotic selectable marker gene or a visible marker gene to allow identification of plant cells transformed with the exogenous DNA sequence. In one embodiment the expression vector further includes a prokaryotic selectable marker gene and a prokaryotic origin of replication that allow for the transformation and reproduction of the expression vector in prokaryotes.

In accordance with the present invention, a DNA construct comprising the regulatory element of SEQ ID NO: 2, a core promoter and a gene operably linked to the core promoter is used to transform a plant cell, using procedures known to those familiar with the art. Such transformation procedures include but are not limited to microinjection, microprojectile bombardment, electroporation, calcium chloride permeabilization, polyethylene glycol permeabilization, protoplast fusion or bacterial mediated mechanisms such as Agrobacterium tumefaciens or Agrobacterium rhizogenes.

Transformed cells (those containing the DNA inserted into the host cell's DNA) are selected from untransformed cells through the use of a selectable

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marker included as part of the introduced DNA sequences. Transformed cells/plant entities can also be identified by the expression of a visible marker included as part of the introduced DNA sequences. Visible markers include genes that impart a visible phenotypic trait such as seed color (i.e., yellow, purple or white genes) or shape (i.e., shrunken or plump genes). Selectable markers include genes that provide antibiotic resistance or herbicide resistance. Cells containing selectable marker genes are capable of surviving in the presence of antibiotic or herbicide concentrations that kill untransformed cells. Examples of selectable marker genes include the *bar* gene which provides resistance to the herbicide Basta, the *npt*II gene which confers kanamycin resistance, and the *hpt* gene which confers hygromycin resistance. An entire plant can be generated from a single transformed plant cell through cell culturing techniques known to those skilled in the art.

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In one embodiment a transgenic plant entity is provided wherein the plant entity consists essentially of a plant cell, seed or plant produced from the in vitro introduction of an exogenous nucleic acid sequence into a plant cell, wherein the exogenous nucleic acid sequence encodes a gene whose expression is controlled by the regulatory elements of SEQ ID NO: 2. More particularly, the transgenic plant is generated by transforming a plant cell with a DNA vector comprising a promoter, having a consecutive 20 base pair sequence identical to the sequence of SEQ ID NO: 2 operably linked to a gene. In one embodiment, the DNA vector used to transform the plant cell comprises the 520 bp sequence of SEQ ID NO: 2 operably linked to a gene. The gene may encode for any product that is beneficial to the plant (for example, gene products that directly or indirectly provide herbicide resistance, insecticidal resistance, fungal resistance or act as growth regulators) or may encode for pharmaceutical or polymer components that are subsequently purified from plant material for commercial use. The exogenous nucleic acid sequences used to produce the transgenic plant typically also include a selectable marker gene or a visible marker gene to allow identification of the cells transformed with the exogenous DNA sequence. In accordance with one embodiment, a plant expression vector comprising a regulatory element operably linked to a non-natively associated gene is used to produce a transgenic plant, wherein the regulatory element is selected from the sequence of SEQ ID NO:2.

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The regulatory element of SEQ ID NO:2 has been demonstrated to be highly efficient in transcribing genes in *Arabidopsis* cells (see Example 2 for details). As shown in Fig. 2 the regulatory element of SEQ ID NO: 2 ligated to the *gusA* coding sequence induced GUS activity at the level of 1.72 nmol MU/hr/1,000 protoplasts, whereas the 35S Cauliflower Mosaic Virus promotor when operably linked to the *gusA* coding sequence produced GUS activity at 1 nmol MU/hr/1,000 protoplasts. The regulatory element of SEQ ID NO: 3 ligated to the *gusA* coding sequence exhibited low level of GUS activity in *Arabidopsis*. Accordingly the 530 bp region, as shown in SEQ ID NO: 2, functions as a strong promoter when operably linked to an exogenous gene in the orientation that naturally expresses the ORF 3 gene in *Arabidopsis*.

Example 1

Isolation of the Genomic Fragment Encoding SEQ ID NO: 1

A genomic library of *Arabidopsis thaliana* ecotype RDL (prepared by ligation of *Hind*III partially digested genomic DNA fragments, ranging between 8-23kb, into the *Hind*III site of the binary cosmid pBIC20) was screened with a *BgI*I fragment of the rice anther-specific cDNA clone RTS-1 (SEQ ID NO: 4) to isolate DNA fragments containing homologous sequences.

The RTS-1 cDNA clone is a tapetum specific gene that encodes an alanine-rich protein that is expressed in tapetum cells of rice anthers. The gene is more fully described in PCT application serial no. PCT/US96/16418, published on April 17, 1997 (publication no. WO97/13401), the disclosure of which is expressly incorporated herein.

Library screening was performed in large Petri dishes (20x20 cm) containing approximately 20,000 recombinant colonies of *E. coli* NM554 cells. Such density should represent about three *Arabidopsis* genome equivalents. The recombinant colonies were lifted on Hybond-N hybridization transfer membranes (Amersham) and membrane-bound DNA (UV irradiation) was probed with the *BglI* cDNA fragment of the RTS-1 gene (SEQ ID NO: 4). Membranes were prehybridized at 50°C for 1 hr in pre-hybridization solution containing 5xSSPE, 5x Denhardt's solution, 0.5% SDS, and 0.2 mg/ml denatured salmon-sperm DNA. Hybridization was

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overnight at 50°C. The filters were washed twice at 50°C in 3 x SSC solution for 15 min, once at 50°C for 15 min in 1 x SSC solution, and in 0.2 x SSC solution at 50°C for 15 min followed by 30 min incubation at room temperature. Washed filters were wrapped in SaranWrap and autoradiography was carried out overnight.

Twenty-three independent clones hybridizing to the probe were identified and selected for endonuclease restriction analysis and Southern blotting. Most of the clones gave rise to multiple signals of varying intensity upon probing with the Bg/I cDNA fragment of the RTS-1 gene (SEQ ID NO: 4). The initial endonuclease restriction and Southern blot analysis identified genomic clone #2 as having a 4.8 kb Hind/III fragment that hybridizes to the RTS-1 probe. This clone was selected for further detailed analysis. When the 4.8 kb Hind/III fragment was subsequently restricted into the two 2.2 kb and 2.4 kb EcoRl fragments, both fragments hybridized to the probe indicating two independent probe binding sites.

The 4.8 kb *Hind*III DNA fragment was sequenced using standard techniques. For sequence analysis, the 2.2 and 2.4 *Eco*Rl fragments internal to the 4.8 kb *Hind*III fragment (See Fig. 1) were subcloned into the pBluescript KS +/-vector. In addition, DNA fragments generated by digestion of the *Eco*Rl fragments with *Xba*I were subcloned to facilitate the sequencing process. The sequence of the cross-hatched region shown in Fig. 1 is shown as SEQ ID NO: 1

A simple homology search for sequences similar to the RTS-1 probe resulted in the identification of three possible binding sites within the 4.8 kb *Hind*III fragment (indicated as boxes above the cross-hatched region of Fig. 1). The matching percentage was in the range of 35-39% over the 190 bp probe fragment. Experimental results on restriction fragment hybridization to the RTS-1 probe were in agreement with predicted positions of the probe binding sites. The longest open reading frame is located in one region of probe binding and it was selected for further analysis.

A genomic fragment from position 825 to 3694 nucleotide SEQ ID NO: 1 was identified as containing sequences homologous to the human splicing factor ASF/SF2 and the *Arabidopsis* SR1 gene, and that region was designated as the ASR-2 region (See Fig. 1). The alignment of genomic DNA sequences of this gene with the SR1 cDNA sequences indicated the presence of eleven putative exons with 82% identity (92% similarity) of the deduced amino acid sequences. The sequence identity

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to the human splicing factor SF2 was 62% as compared to 59% identity between the SR1 and SF2 genes. An identical structural organization of RNA-binding domains, the glycine spacer, and the SR domain was observed among all three genes. The ASR-2 coding sequence also included a highly charged PSK domain at the C-terminal end similar to the SR1 gene but absent in the ASF/SF-2 gene coding sequences.

The presence of ASR-2 transcripts was analyzed by reverse transcription PCR in different parts of *Arabidopsis* plant. Total RNA was isolated from various *Arabidopsis* organs and reverse transcription of 5 μ g of the total RNA, treated with RNase-free DNase, was performed with MuMLV-reverse transcriptase (400 units) and oligo dT₁₈₋₂₂ primer (4 μ g) for 1 h at 42 °C followed by 5 min at 95 °C. Following incubation, the reaction mixture was treated with RNAse H (8 units) for 20 min at 37 °C. Five μ l of the reverse transcription reaction was then amplified with Taq-polymerase (Perkin-Elmer Cetus) using primers that recognize the first exon in the RNA recognition domain and the SR domain of ASR-2. Primer sequences were selected that were specific to the ASR-2 domains but not to the SR1 homologous domains.

The ASR-2 gene was found to be expressed in all plant parts investigated including the leaves, stems, siliques, and roots. Similar levels of expression were observed in different plant organs. The experiment revealed the presence of more than one transcript hybridizing to the ASR-2 probe (the 2.4 kb EcoRI fragment of ASR-2 genomic clone). Shorter transcripts were identified in RT-PCR reaction products than expected and could represent splice variants of ASR-2 transcripts. Sequencing of the amplified major RT-PCR product confirmed all predicted intron-exon junction sites except the splicing sites (5' as well as 3') of the intron #7. Such transcripts contain the SR domain message but they cannot be translated into the full length protein because splicing of the intron #7 generates a frame shift mutation leading to a stop codon just after the splice site.

Example 2

30 Expression of gus A using the promoters of SEQ ID NO: 2 and SEQ ID NO: 3

Arabidopsis protoplasts were transformed with bacterial vectors
containing the 5' untranslated ASR-2 DNA sequence connected to the coding

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sequences of the bacterial b-glucuronidase gene. The coding sequences were ligated to the 3' end and to the 5' end of the promoter sequence and the respective constructs were designated as pUN-GUS [having the promoter orientated in the direction that normally transcribes the ORF-3 gene (i.e., SEQ ID NO: 2) and operably linked to the gusA gene] and pASR-GUS [having the promoter orientated in the direction that normally transcribes the ASR-2 gene (i.e., SEQ ID NO: 3) and operably linked to the gusA gene]. The sequences of the junction site between the promoter sequence and the gusA coding sequence are disclosed as SEQ ID NO: 5 for pASR-GUS and SEQ ID NO: 6 for pUN-GUS wherein the ATG start codon is located at nucleotide 6 and the coding region of the gusA is located at nucleotide 30.

The unique DNA promoter sequence discovered and claimed in the present invention is located between the nucleotides at the position 3690 through 4221, of SEQ ID NO: 1. The sequence is 530 nucleotides in length and is presented as SEQ ID NO: 2 and SEQ ID NO: 3. The 530 bp sequence contains numerous transcription factor binding sites including two "TATA" boxes at positions 141, 316 and "CAAT" boxes located at 223, 386, 448, and 486 and one zeste element (GTGAGTG) at 264 of SEQ ID NO: 2.

The activity of the claimed sequence driving the expression of a foreign gene, gusA, in plant cells was compared to the activity of the 35S cauliflower mosaic 20 virus promotor in Arabidopsis protoplasts. Four expression vectors p35SGUS (having the 35S cauliflower mosaic virus promotor operably linked to the gusA gene), pUN-GUS (as described above), and pASR-GUS (as described above) and a control vector lacking a promoter operably linked to the gusA gene (DNA-), were introduced into the protoplasts by a PEG-mediated transformation procedure. One day after 25 transformation, the GUS activity was determined. The 35S CaMV promotor controlling the gusA sequence produced GUS activity at 1 nmol MU/hr/1,000 protoplasts, while the claimed sequence ligated to the gusA coding through its 3' end (SEQ ID NO: 2) induced GUS activity at the level of 1.72 nmol MU/hr/1,000 protoplasts (see Fig. 2). The claimed sequence ligated to the gusA coding sequence in 30 the opposite orientation (SEQ ID NO: 3) exhibited a low level of GUS activity in Arabidopsis.

-11-

SEQUENCE LISTING

5	(1) GENE	RAL INFORMATION:
J	(i)	APPLICANT: Purdue Research Foundation, Hodges, Thomas K. Lysnik, Leszek A
10	(ii)	TITLE OF INVENTION: Regulatory Element For Expressing Genes In Plants
	(iii)	NUMBER OF SEQUENCES: 6
15	(iv)	CORRESPONDENCE ADDRESS: (A) ADDRESSEE: Barnes & Thornburg (B) STREET: 11 S. Meridian (C) CITY: Indianapolis (D) STATE: Indiana
20		(E) COUNTRY: USA (F) ZIP: 46204
25	(v)	COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
30	(vi)	CURRENT APPLICATION DATA: (A) APPLICATION NUMBER: (B) FILING DATE: (C) CLASSIFICATION:
35	(viii)	ATTORNEY/AGENT INFORMATION: (A) NAME: Breen, John P. (B) REGISTRATION NUMBER: 38,833 (C) REFERENCE/DOCKET NUMBER: 3220-29933
40	(ix)	TELECOMMUNICATION INFORMATION: (A) TELEPHONE: (317) 231-7745 (B) TELEFAX: (317) 231-7433
45	(2) INFO	RMATION FOR SEQ ID NO:1:
	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 5285 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double
50		(D) TOPOLOGY: linear
	(ii)	MOLECULE TYPE: DNA (genomic)
55	(iii)	HYPOTHETICAL: NO
	(iv)	ANTI-SENSE: NO
60	(vi)	ORIGINAL SOURCE: (A) ORGANISM: Arabidopsis thaliana
	(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:1:
65	GAATTCCAC	GC GTGGAAGAGA CCAGGACAAC AAACAGCGAG TTTGTATAAA GAAGCCCAAC 60
.		GG AGTAAGAGAC GAATCCGCCG CGGTGGATGA AGAAGCGGAT GCGGCGGCGG 120
	AGGGAGGAI	AG AGGGAGGAGA TCGGAGATGA CGGTGGGGAT AGCGGAGTAG CACTGAGTTT 180

	TACAACAAAC	GTTTCGAACA	ATTGAGGAGA	ATTTGGTGCA	GACGCAAGCG	AGACGAGCCC	240
	AGTTTCTTGG	ATCATCTTGG	AGCTTGAAGA	AGATGTTGAA	GACTACGTCT	TCTGGTATAC	300
5	AAGAGAAGAC	AGATTCCGCC	ATGGATCGCC	TTCTTCTCTC	TAGGCGGCTC	CTTTCTTATC	360
	CAAATTCACT	TATACTGTTA	TGGGTCCGGT	ACGCGTAAAC	CGGGAATAGT	CTTAACTGTT	420
10	CTTAAGGTGG	GTCACAGATT	CACTAACACC	CACACAAAGG	CAAGTAAGTA	ATGCGCAACA	480
10	GCTCTCGAAA	ATGACATCGT	ACGGACTGAA	CTAAAATGTA	AAGGGTCCGG	GTATCAAAAT	540
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15	TCGAATTGTG	AGCTTTTGAA	TTAAGTTTGG	TATTCGACAG	TAATTTTTGA	TAGTTCGTTT	660
	TAAGCACTAA	CTATATTAGC	AAGTCATATA	AATCAGCTGA	GCTTAGCTCA	TAAACTGATG	720
20	ATGACTGAGT	ATATATCATT	CCCATGTGCA	AACCCAAGCT	AATAAGAATG	AAATACACAA	780
	CTAGTTTTTC	AACTTCTCAT	ACATAAGAGA	GATCATCTTT	ATGAGAATCT	TCCAACAGAC	840
	CCAGCTCTTT	TACCTGATAT	GAGATGTTTC	TTTACCTGTA	AAACAGTTGG	AAAGAGATTC	900
25	AGAGAAAGAG	TGAGAATGTT	CCAAGCAAGT	TAAAAAGAGT	GTGCATTACC	GAGATGGACT	960
	CCTGCTTCTG	CTCTTGCTCC	TACCACGGAT	AGGGCTCAGC	TGCTTGCTAG	GGCTCTTACT	1020
30	TGCTTCCTTC	TGAACCTGCA	ACACACACAA	TTCAATCCAG	ATGATGAGAA	TGTAATTAGC	1080
	TAGACATAGA	TCATGTCTTT	TGGATAGTAT	GGATTGAAAA	CCGAAAATAT	TGTGCTGATA	1140
	GTATAGCGGA	GTAAAAAGTG	TATTAGAGAT	AGATGACTTA	GAGAGGGTAG	AGGAGATCTT	1200
35	GACCTTGAAC	GAGATCTGCA	AAAGTCCGAG	AAACAAATCC	AGTTTTAAAA	ATCCAATATT	1260
	TCCGTGTTTA	ATAGCTGTTT	CACCAAGACC	CTTGATGCCT	CATTCCATGT	CAGACATCAA	1320
4 0	AACACAATCA	CAAGAAGCCA	AAAAGGAAAA	AACATATCTT	CAGACTATTT	TTAGCAGAAA	1380
	ATCAAACCAA	CACAAGCCTG	CTTTTTGTTA	AAACAGATGG	TAGAGATGGA	GAATATCAAA	1440
	GCAAATCTAA	TTTTATCAAA	CCCTGTGTCC	AGAACAGCAT	CGGTTCCATG	AGAATCCAGA	1500
15	TCGCTTGTCG	ATTAGCAGGT	AATGAAAATA	TCTGTCCTAG	CTCGATGGGT	CCATTTTGAT	1560
	GACACTATAT	CAATGCGATC	CAATGTCTCT	CCACTGTTAC	CCATTTAGCA	GCAATGGATA	1620
50	TTATCAGAAA	ACGAACTTGC	CCATTTAAGA	AAGAGCATAT	ACCTTTGAGA	CGAAACAGTT	1680
-	GTGTACAACC	AACCAAGACA	TTCCATGATC	CAGATACTAT	TTCCCATATT	TTAGTTGATT	1740
	GTATGTATAT	CCATATCTAA	GAAACAAAAC,	CATTCTCAAC	ACTATAATTA	TAAAAGACCA	1800
55	GACTTTCAAA	GGAAATAAAT	CGTGTACCCT	TAACTAGAAG	CAATCATCAT	TCTCAATCAA	1860
	CAACTCGGAG	TCATCCCAGT	GACATCTTTT	AATGTGATGT	CACCAAACTT	CAAGGGAAGA	1920
50	GCTAAATAGA	GCCTATGTTA	TGTTTTATGT	TGGATATTTT	AGCATAAACA	TTATAGAAGA	1980
	AATGAAGCAT	ACCCTCGTGG	AGACAGTGAC	CTCGACTTAG	AGCGGGAGCG	agagcgagga	2040
	GATCTCGATG	TAGATTTTGC	AGGCGATCTA	CGCAAAGATT	TAGCCTTTGG	ACTTCTGCTC	2100
55	TTGCTCCTGC	TTCTGCTGCG	GCTACGACTA	CGGCTGGGAC	TCCGTCCACG	GCTGCGGCTC	2160
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	ACCTGTAATG	ACATAGGGAA	ATGTTAAGTG	CAATAAAAAC	GAATGCAGCT	CTCAGAATTC	2280
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5	GAGCTTTTTT	TATCTGGACA	AAATAAAGAA	TATAATCATG	AGTAATCAAG	GATGAACACA	2400
	TTCTCAGCCA	CAGTCCCACA	CAGAAAATCA	AATAATATGA	AGAGGAAGAA	AAACATCTCA	2460
10	CCGCATATTT	CATGTCCTCG	TAGCTGGTAT	AATCTACAAT	TCCAGTTGTA	CCTGTAAATA	2520
10	AATAGTCCAC	GATATAGATT	TTTTAAAGCA	TGCACAGACT	AGTTAAAACA	GGATTTAAAG	2580
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30	CACACATAAA	TTATGTTTGG	CTACCAGTCA	ACAATACAAA	GTTTGTGTAA	AAATTCTGAA	3120
	ATTTGATGAT	TAACAAACCC	GTAAATGATG	CCCATCAAAG	TCATAACCAT	CACGGCCATA	3180
	AATTGCATCA	TCAGCATCAC	GAGCATCCTC	AAACTAAATC	ACATATATCA	CAAAACATTA	3240
35	GTGGTAGTAT	CTCCCAACAT	TTGAAAACTC	ATGAACACTC	AACAACAACG	AAGAGCCTAA	3300
	CTGTAAAATC	AACAACAAGC	CTTTATAAAA	CATGTGGTTG	CATAAAAAAT	CTGACCTCGA	3360
40	CGAATGCATA	GCCTGGAGGC	CTCGGCGGAA	TCTTCAAATC	GATTTGAACA	ACAGGTCCAT	3420
	ACTTCAAAAA	Aaataaggaa	GAACAATTCT	TAAGGAAACT	TCTTCCAAAT	AAAATCAGAA	3480
	TCCAACAATT	CCGAAGATGC	ATTCAATTTC	TTTCACATGC	AAATCTTGTA	AAGACATATT	3540
45	CATCACATAA	CACAAAATTC	GATCCTGAGT	TCTGAGTTCT	TAAATTAGGA	GAAACGAGTA	3600
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50	GACGTAAATC	GTTCTACTCG	AACGGCTGCT	CATTTATTTC	TTTCCTATAC	СААААТСААА	3720
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55	CTAAAATTCC	AGTAACGATT	CCGATCACCT	GAGAGAAAAT	TCCGATGGAA	GAGAAGAAGA	3900
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60	ACGTGACCGA	CCGGGTACGT	ATTAAAATAC	ATTGTGTCTT	GACCGTATAA	AATACATTTG	4020
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	CCAAATCTTC	AATACTTGAT	ATTTCTCAAA	GAACTTGAAA	ACAACACAGA	TCCATTCCCA	4140
65	ATTTCAGATT	CACTCAAAAA	GGATTTTTCT	TTTTTCATTT	TCGCTTTTTG	TGATCTGGAA	4200
	AGTTGTTACC	TTTAACAATG	TCTCCGAAAC	ATCTAGAGTC	ATCACGAAGC	TCTATTGAAT	4260

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	CATGCACTTC ACAGCTTCTC TCATGGCGAC CATTTCACCG CTCCAAAACC CTAGACTCAT	4320
	CTGACCAACC ACCGCAGACC AATGGGTTTC ACTCCTTTAC TCCCAAACGC CCTTGCTTCT	4380
5	CCGATCGATC CACTTCTTTC ACCATCGAAG CTATGAGCCG TCTCTCACTC GCCGACGACG	4440
	ACAATGGAGG GAAGACATTA TCAGCTTCCA ATTACAGCAA TAGAGGAAGT TTCAGGTTAG	4500
10	TAGCGAGGAA GCGGCGGCGG CGTAATTCGA GATCGGTGTC TGGTCGGAGT AGTGATCGGA	4560
	GTGGGACTCG GAGATGTTGC TCCATTGGTG CTCATGGGAC TTGTTCGGAT TTGCCTTTCG	4620
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15	GTGAGGCGGC GAGGAATTCA CGGAGAGAGC GGCGAGATTC TGGTGGAGAG AAGGAAGCTT	4740
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	TGTGTTCCAT CTTGAGATTC GTGGTTTTTG CTATGAAGAT TTGGAGATTG ATCATCATTG	4980
25	ATTAGATTAA AGATGACAAC TTTAGTGTTA TTTCTTCTGA TGAAAATGAG TCTGATTTTG	5040
	CTCTGCTTGT CTATTATGGC ATTGCCTCAT AGGAATTGTC AGAAAGTTGT CAAATTTTGA	5100
30	TATGTTTAGT GATTGGTGAG TGTTTTGGAT GGAATTGGGT TCTTATCATG TTAGGTCATT	5160
	GTCTGAAATG GATATGTATG TACTTGGTAT TTTGATATGT TTAGTGATTG GTGAGTGTTT	5220
	TGGATTTGGA GCAGATACAG ATTCCACAAT GGGGATGTCT GGTGAGACAA ATCTCAGATA	5280
35	GTAAA	5285
-		0200
-	(2) INFORMATION FOR SEQ ID NO:2:	
40	(2) INFORMATION FOR SEQ ID NO:2: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double	
40	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
40	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic)	
40 45	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (iii) HYPOTHETICAL: NO	
40 45	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO	60
40 45	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:	
40 45 50	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: CATTTATTC TTTCCTATAC CAAAATCAAA ATTGAGATTC GAAACGTCAA TAGATCGAAA	60
40 45	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: CATTTATTC TTTCCTATAC CAAAATCAAA ATTGAGATTC GAAACGTCAA TAGATCGAAA CAAAGAAGCG ATCACACACA AAAAAAACTC ATTGGATAAC GATTAACCTA AGGAAAACTA	60 120
40 45 50	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2: CATTTATTC TTTCCTATAC CAAAATCAAA ATTGAGATTC GAAACGTCAA TAGATCGAAA CAAAGAAGCG ATCACACACA AAAAAAACTC ATTGGATAAC GATTAACCTA AGGAAAACTA AAGAGGTTTG ATTGATCGTC TATATATGAA CTAAAATTCC AGTAACGATT CCGATCACCT	60 120 180
40 45 50	(i) SEQUENCE CHARACTERISTICS:	60 120 180 240
40 45 50 55	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 530 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO (**i) SEQUENCE DESCRIPTION: SEQ ID NO:2: CATTTATTC TTTCCTATAC CAAAATCAAA ATTGAGATTC GAAACGTCAA TAGATCGAAA CAAAGAAGCG ATCACACAC AAAAAAACTC ATTGGATAAC GATTAACCTA AGGAAAACTA AAGAGGTTTG ATTGATCGTC TATATATGAA CTAAAATTCC AGTAACGATT CCGATCACCT GAGAGAAAAT TCCGATGGAA GAGAAGAAGA AAGGCGAAAA TTGAAATCTG ACTAGGGCTT TCGAATACCA TAGAGATCAT CACGTGAGTC ACGTGACCGA CCGGGTACGT ATTAAAATAC	60 120 180 240 300

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10	(C) STRANDEDNESS: double (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
15	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: NO	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:	
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25	ATCAAGTATT GAAGATTTGG AATTGTTCTT TTGGGTTTAA GAGCTTTTGA TTGAAGATTA	180
	CGATTTGTTG CAAAACGGGT CAAATGTATT TTATACGGTC AAGACACAAT GTATTTTAAT	240
30	ACGTACCCGG TCGGTCACGT GACTCACGTG ATGATCTCTA TGGTATTCGA AAGCCCTAGT	300
	CAGATTTCAA TTTTCGCCTT TCTTCTTCTC TTCCATCGGA ATTTTCTCTC AGGTGATCGG	360
	AATCGTTACT GGAATTTTAG TTCATATATA GACGATCAAT CAAACCTCTT TAGTTTTCCT	420
35	TAGGTTAATC GTTATCCAAT GAGTTTTTTT TGTGTGTGAT CGCTTCTTTG TTTCGATCTA	480
	TTGACGTTTC GAATCTCAAT TTTGATTTTG GTATAGGAAA GAAATAAATG	530
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50	(ii) MOLECULE TYPE: DNA (genomic)	
30	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: NO	
55	<pre>(vi) ORIGINAL SOURCE: (A) ORGANISM: Oryza sativa</pre>	
60	(vii) IMMEDIATE SOURCE: (B) CLONE: RTS-1	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
65	GAGCCGCCCA CCGATGACGG CGCGGTCCGG GTGGCGGCGG GGCTGACGAA GTGCGTGTCC	60
	GGGTGCGGTA GCAAGGTGAC CTCCTGCTTG CTCGGCTGCT ACGGCGGCGG CGGCGGCGCC	120
	GCCGCCGCCG CGACGGCGAT GCCGTTCTGC GTCATCGGCT GCACCAGCGA CGTCTTGTCC	180

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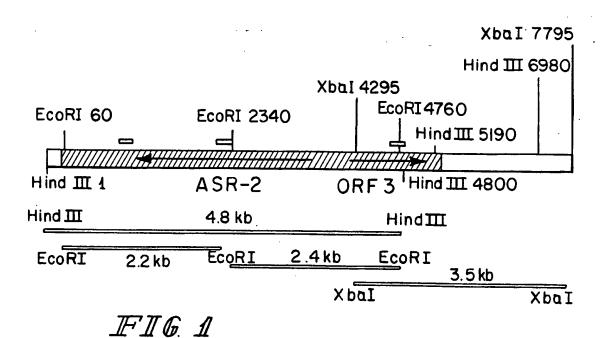
	TGCGCC	186
5	(2) INFORMATION FOR SEQ ID NO:5:	
10	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 38 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
	(ii) MOLECULE TYPE: DNA (genomic)	
15	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: NO	
20		
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
	AATAAATGAG CCCGGGTGGT CAGTCCCTTA TGTTACGT	38
25	(2) INFORMATION FOR SEQ ID NO:6:	
30	 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 38 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear 	
35	(ii) MOLECULE TYPE: DNA (genomic)	
,,	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: NO	
40	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:	
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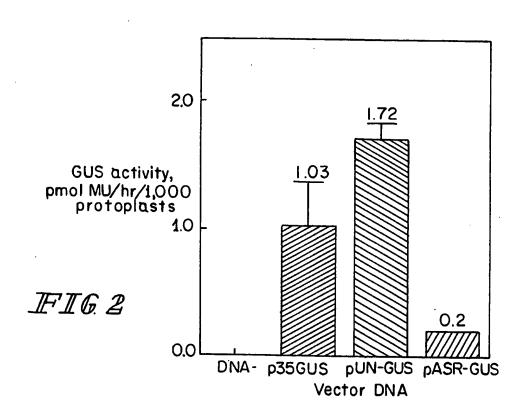
CLAIMS:

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- 1. A substantially pure nucleic acid sequence comprising a sequence as set forth in SEQ ID NO: 2.
- 5 2. A DNA sequence comprising a 20 base pair nucleotide portion identical in sequence to a consecutive 20 base pair portion of the sequence as set forth in SEQ ID NO:2.
 - 3. A recombinant expression vector comprising the DNA sequence of claim 2 operably linked to a gene.
 - 4. The expression vector of claim 3, further comprising a gene encoding a eukaryotic selectable marker.
 - 5. The expression vector of claim 4, further comprising nucleic acid sequences that enable replication of the expression vector in a bacterial host, and a gene encoding a bacterial selectable marker.
- 6. A plant entity consisting essentially of a plant cell, seed or plant produced from the *in vitro* introduction of an exogenous nucleic acid sequence of claim 3.
 - 7. An expression vector comprising the DNA sequence of claim 2 and a polylinker sequence.
 - 8. The expression vector of claim 7, further comprising a gene encoding a eukaryotic selectable marker.
 - 9. The expression vector of claim 8, further comprising nucleic acid sequences that enable replication of the expression vector in a bacterial host, and a gene encoding a bacterial selectable marker.
- 25 10. A transgenic plant entity comprising plant cells transformed with a DNA sequence comprising a regulatory element operably linked to an exogenous gene, said regulatory element comprising a 20 base pair nucleotide portion identical in sequence to a consecutive 20 base pair portion of the sequence as set forth in Seq ID NO:2.

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INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/06761

A. CLASSIFICATION OF SUBJECT MATTER IPC(6) :A01H 5/00; C07H 21/04; C12N 15/63, 15/82						
US CL :435/320.1; 536/24.1; 800/298						
According to International Patent Classification (IPC) or to both national classification and IPC						
B. FIELDS SEARCHED						
Minimum documentation searched (classification system followed by classification symbols)						
U.S. : 435/320.1; 536/24.1; 800/298						
Documentation searched other than minimum documentation to	the extent that such documents are included in the fields searched					
Electronic data base consulted during the international search (name of data have and where practicable search terms used)					
Picase See Extra Sheet.	version of call outs and, whose placeholder, scales with used)					
C. DOCUMENTS CONSIDERED TO BE RELEVANT						
Category* Citation of document, with indication, where	appropriate, of the relevant passages Relevant to claim No.					
A FISSCHER et al. Identification of Pothe Far-Upstream Region of the Ara Promoter. Plant Molecular Biology. 3, pages 873-886, see entire document	bidopsis thaliana Plastocyanin November 1994, Vol. 26, No.					
DOLFERUS et al. Differential Interactions of Promoter Elements in Stress Responses of the Arabidopsis Adh gene. Plant Physiology. August 1994, Vol. 105, No. 4, pages 1075-1087, see entire document.						
A LAZAR et al. Identification of a Plan Similar to the Mammalian Splicing I Acad. Sci. USA. August 1995, Voentire document.	Factor SF2/ASF. Proc. Natl.					
X Further documents are listed in the continuation of Box (C. See patent family annex.					
Special categories of cited documents:	"T" later document published after the international filing date or priority					
"A" document defining the general state of the art which is not considered to be of particular relevance	date and not in conflict with the application but cited to understand the principle or theory underlying the invention					
E earlier document published on or after the international filing date	"X" document of particular relevance; the claimed invention cannot be					
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other	considered novel or cannot be considered to involve an inventive stap when the document is taken alone					
special reason (as specified)	'Y' document of particular relevance; the claimed invention cannot be					
O° document referring to an oral disclosure, use, exhibition or other combined with one or more other such documents, such combination being obvious to a person skilled in the art						
P° document published prior to the international filing date but later than "&" document member of the same patent family						
Date of the actual completion of the international search	Date of mailing of the international search report					
29 JULY 1998	18 AUG 1998					
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks	Authorized officer					
Box PCT	THANDA WAI					
Washington, D.C. 20231 THANDA WAI Csimile No. (703) 305-3230 Telephone No. (703) 308-0196						

INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/06761

		FC1/0398/00/0	
C (Continua	tion). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.
Á	LOPATO et al. Characterization of a Novel Arginine/Serine-Rich Splicing Factor in Arabidopsis. Plant Cell. December 1996, Vol. 8, No. 12, pages 2255-2264, see entire document.		1-10
A,P	SU et al. Arabidopsis ASF-SF2 Transcripts are Alterna Spliced. Plant Physiology. July 1997, Vol. 114, No. 3 Supplement, page 246, see Abstract No. 1260.	atively ,	1-10
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INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/06761

B. FIELDS SEARCHED Electronic data bases consulted (Name of data base and where practicable terms used):						
APS, MEDLINE, AGRICOLA, BIOSIS, EMBASE, WPIDS						
search terms: ASF/SF2, ASR-2, SR1, arabidopsis, splic? factor#, regulator? element#, express?, promotor#, promotor#						
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